Mini Review

The Role of Endothelial Cells in the Resolution of Acute Inflammation

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ABSTRACT

Endothelial cells are key regulators of the inflammatory response. Lining blood vessels, they provide in the steady state an antiinflammatory, anticoagulatory surface. However, in the case of injury or infection, endothelial cells control the adhesion and migration of inflammatory cells, as well as the exchange of fluid from the bloodstream into the damaged tissue. Thus, expression of endothelial adhesion molecules, cytokines, and changes in permeability need to be tightly regulated to allow for a controlled inflammatory response. Acute inflammation is characterized by tissue infiltration of neutrophils, followed by monocytes/macrophages. For successful tissue regeneration and healing, the acute inflammatory response needs to be actively shut down, a process called resolution of inflammation. Unsuccessful resolution may lead to excessive tissue damage and ultimately results in chronic, self-promoting inflammation. This review will summarize recent advances in the field of endothelial biology, which point to an active participation of the endothelial barrier in the resolving process. *Antioxid. Redox Signal.* 7, 1744–1754.

INTRODUCTION

SELF-LIMITING ACUTE INFLAMMATION caused by injury or infection is characterized by early vascular leakage, with extravasation of plasma components and fluid, and massive recruitment of neutrophils that ingest invading pathogens and release proinflammatory mediators and reactive oxygen species (ROS) leading to tissue damage, and oxidation of membrane lipids; subsequently, neutrophils undergo apoptosis, an important step in the resolution of inflammation (for reviews, see 82, 83). As a consequence, specific monocyte infiltration and maturation into macrophages are initiated to remove apoptotic cells, fibrin and protein clots, and bacterial and cellular debris; additionally, proinflammatory mediators are dissipated and antiinflammatory, proresolving mechanisms are induced; finally, normal vascular permeability has to be restored, ultimately leading to a cessation of leukocyte emigration and restoration of normal tissue function.

A continuous imbalance between initiation of proinflammatory mechanisms and those that promote resolution leads to prolongation of an inflammatory response. Thus, chronic inflammation may arise from failure and inefficiency of mechanisms normally responsible for resolution of inflammation and restitution of tissue homeostasis. Although much has been learned about the proinflammatory pathways over the last years, mechanisms that lead to the equally important resolution of acute inflammation are less well understood. Recent evidence demonstrates that resolution is an active process involving apoptosis of neutrophils, selective attraction and infiltration of monocytes and their differentiation into macrophages and dendritic cells, and up-regulation of proresolving genes such as heme-oxygenase-1 (HO-1) and cyclooxygenase-2 (COX-2) (109). Hence, the mechanisms involved in the endogenous inflammation-resolving process are currently extensively investigated, as they could offer possible targets in the treatment of chronic inflammation (89, 91, 109).

ENDOTHELIAL CONTROL OF LEUKOCYTE ADHESION AND BARRIER FUNCTION

The endothelial cell layer forms a barrier between blood and tissue. At sites of acute inflammation, leukocytes have to pass this barrier in order to exert their function toward invading microbes. During resolution of acute inflammation, the physiological barrier has to be restored to limit cell and fluid extravasation. Therefore, the endothelium plays a key role not only in promoting, but also ending inflammatory responses after injury or infection (Fig. 1).

Endothelial cells can actively regulate the migration of leukocytes through the vessel wall by expressing adhesion molecules and chemokines (67) on their luminal surface, allowing leukocytes to adhere and finally migrate into the tissue. For successful resolution, endothelial cells must first selectively let pass mononuclear cells for the clearance of

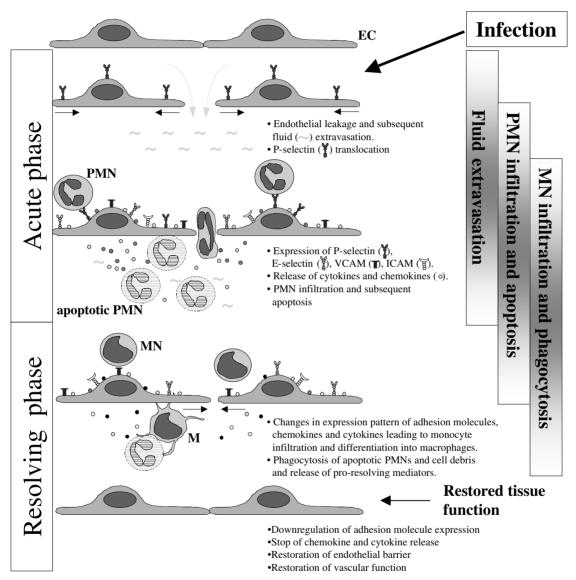


FIG. 1. The role of the endothelial cell in the resolution of acute inflammation. Upon inflammatory stimulation, the endothelial barrier function is rapidly lost, leading to extravasation of fluid. Preformed P-selectin is translocated to the luminal surface of endothelial cells, adhesion molecules such as E-selectin, ICAM-1, and VCAM-1 are expressed, and chemokines are expressed and released. These changes lead to transmigration of neutrophils through the endothelium into the tissue. Eventually, neutrophils undergo apoptosis. During this phase, more and more monocytes are selectively recruited to the site of inflammation, presumably by changes in the expression pattern of adhesion molecules and chemokines on the endothelial cell. The accumulating monocytes differentiate into macrophages to phagocytose apoptotic neutrophils and cell debris. Finally, release of proinflammatory chemokines is abrogated and endothelial barrier and vascular function is restored. During the whole process of acute, resolving inflammation, the endothelial cell serves as a gatekeeper, controlling which cell type enters the site of inflammation. EC, endothelial cell; PMN, polymorphonuclear leukocyte; Mφ, macrophage.

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cellular corpses, whereas the drainage of extravasated fluids is facilitated by the lymphatic system at the site of inflammation. Furthermore, proinflammatory chemokines have to be dissipated. In this context, the importance of the chemokine receptor D6 in resolution was recently illustrated. It was shown that D6, which is mainly expressed on lymphatic endothelial cells, binds, internalizes, and subsequently degrades a variety of chemokines and thus promotes resolution of cutaneous inflammation (44). Finally, endothelial cells have to actively shut down the expression of adhesion molecules (Fig. 1). Recent data demonstrate that neutrophils by themselves down-regulate adhesion molecule expression in endothelial cells and reduce binding of inflammatory cells (96). On the other hand, in vitro experiments showed that angiopoietin-1 (Ang-1) down-regulates expression of E-selectin (26) and of thrombin-induced interleukin-8 (IL-8), and thus inhibits neutrophil binding to endothelial cells (78). Furthermore, overexpression of Ang-1 in mice abrogated lipopolysaccharide (LPS)-induced up-regulation of E-selectin, intercellular adhesion molecule-1 (ICAM-1), and vascular cell adhesion molecule-1 (VCAM-1) and preserved normal endothelial nitric oxide synthase (eNOS) expression (110), significantly improving survival of mice challenged with lethal doses of LPS. Moreover, vessels of mice overexpressing Ang-1 exhibited resistance to vascular leakage (100, 101), implicating Ang-1 as an interesting target in promoting resolution.

Superoxide dismutase (SOD) and ROS scavenging: Influence on redox signaling

Many proinflammatory intracellular signaling pathways in endothelial cells critically depend on free radicals. Whereas in granulocytes ROS generation by NADPH oxidase is necessary for successful antimicrobial defense during acute inflammation, activation of this enzyme in endothelial cells has been implicated in the progression of endothelial dysfunction and atherosclerosis (81). Therefore, inhibiting ongoing inflammatory responses by reducing oxidative stress is another potential mechanism to promote resolution. Copper, zinc superoxide dismutase (CuZn-SOD) scavenges superoxide radicals by production of hydrogen peroxide and oxygen. A recent study shows that mice overexpressing CuZn-SOD were protected against LPS-induced endothelial dysfunction (20). Furthermore, endothelial cells overexpressing SOD showed upon tumor necrosis factor- α (TNF α) stimulation decreased superoxide production, decreased surface expression of VCAM-1 and ICAM-1, and thus decreased neutrophil adhesion. Moreover, in these cells, TNFα-induced activation of activator protein-1 (AP-1) and nuclear factor-κB (NFκB) were significantly reduced (63). Besides SOD, selenoproteins such as glutathione peroxidases and thioredoxin reductases were shown to prevent oxidative events in endothelial cells (for review, see 8). Whether these enzymes are up-regulated during a self-resolving inflammation and whether their induction would support resolution remain to be investigated.

Leukocyte specific protein 1 (LSP-1)

LSP-1 was described to be a substrate for protein kinase C and for mitogen-activated protein kinase-activating protein (MAPKAP) kinase, and to play an important role in the regu-

lation of cytoskeletal architecture and motility. Initially, LSP-1 was thought to be restricted to leukocytes; however, recently LSP-1 was shown to be present also in endothelial cells. Surprisingly, using chimeric mice that lack LSP-1 either in endothelial cells or in leukocytes, it was clearly shown that endothelial, but not leukocyte LSP-1 controlled the process of leukocyte transmigration toward TNF α (64). By using a zymosan-induced peritonitis model (45) or a zymosan-induced knee inflammation model (106), it was shown that mice that lack LSP-1 exhibited an increase in neutrophil accumulation. Whereas rolling and adhesion of leukocytes remained unaltered, extravasation was clearly increased in LSP-1-deficient mice. Therefore, LSP-1 displays a new target in endothelial cells that could contribute to the resolution of inflammation.

Endothelial CD39 and adenosine release

CD39, an ecto-nucleoside triphosphate diphosphohydrolase (NTPDase), which is expressed on quiescent vascular endothelial cells and can be induced by hypoxia (23), efficiently hydrolyzes extracellular ATP and ADP to AMP and adenosine. Mice lacking CD39 showed exacerbated inflammatory responses to chemical skin irritants, due to their inability to hydrolyze extracellular accumulating ATP (69). Furthermore, it was shown that ATP released from activated neutrophils during transcytosis is metabolized by endothelial CD39, especially under hypoxic conditions. The liberated adenosine leads via activation of its receptor (AdoRA2B) to an increase in endothelial cylic AMP, which finally leads to enhanced barrier function and, in in vivo models, to decreased vascular leakage (23; 35). In vitro, adenosine attenuates polymorphonuclear leukocyte (PMN) transendothelial migration (105). Moreover, it was shown that endothelial cells themselves release ATP upon LPS stimulation that precedes IL-1 α secretion. When those endothelial cells overexpressed CD39, secretion of IL-1α was abrogated (41). Additionally, endothelial CD39 expression inhibited ATP-induced apoptosis and NFkB activation and abrogated E-selectin expression (33). Therefore, induction of CD39 and thus increased adenosine liberation followed by improved barrier function would result in a stop of leukocyte migration during the resolving phase of inflammation.

AOC3 (VAP1): oxidative modification of adhesion molecules

Recently, it was suggested that leukocyte transmigration is dependent on the enzymatic activity of endothelial amine oxidase, copper-containing-3 (AOC3), also known as vascular adhesion molecule 1 (VAP1) (53). Moreover, *in vivo* inflammation studies clearly showed that when AOC3 activity is lost either by treating animals with antibodies against AOC3 or by using mice lacking the enzyme, leukocyte transmigration through the endothelium is significantly decreased, whereas the rolling velocity of neutrophils is greatly increased (95, 102). Whether endothelial AOC3 is actively shut down, as well as the mechanism behind it, still remains unclear; however, regulating the function of this enzyme may be important in promoting resolution of inflammation.

ENDOTHELIUM-DERIVED GASEOUS MEDIATORS IN THE RESOLUTION OF ACUTE INFLAMMATION

HO-1

Heme exerts several proinflammatory actions, such as upregulation of cytokines and chemokines, and inhibition of neutrophil apoptosis (3). Therefore, degradation of heme is important for tissue homeostasis. HO-1 is the rate-limiting enzyme of heme catabolism, catalyzing the breakdown of heme into carbon monoxide (CO), iron, and bilverdin, which is further metabolized to bilirubin. It is well established that expression of HO-1 is cytoprotective in a variety of cell types, including endothelial cells. Moreover, HO-1 is highly up-regulated during the resolution of inflammation (107, 108), and inhibition of HO-1 leads to increased inflammatory cell extravasation and prolongation and potentiation of inflammation. Therefore, a central role of HO-1 and its products in the resolution of inflammation, but also in wound healing and angiogenesis, was suggested.

In vivo experiments clearly demonstrated that induction of HO-1 inhibited oxidant-induced (37) and shock-induced (70) leukocyte rolling and adhesion. HO-1 affected expression of P-selectin on the endothelium, whereas expression of adhesion molecules on leukocytes remained unchanged (37), again pointing to an important role of the endothelium in control of leukocyte migration during resolution. Indeed, overexpression of HO-1 in endothelial cells attenuated TNF α -induced NF α B activation and protected against proinflammatory responses, including increased VCAM-1, E-selectin, and monocyte chemotactic protein-1 expression and decreased eNOS expression (52). Interestingly, these antiinflammatory and potentially proresolving effects were mediated by HO-1-derived bilirubin, but not by CO (94).

HO-1-derived CO has been described to be beneficial in a variety of inflammatory models (for review, see 75). For instance, CO suppresses LPS-induced TNF α production in leukocytes *in vivo* and *in vitro* (71, 76). However, on endothelial cells, CO seemed to mediate mainly antiapoptotic effects (93). For example, it was shown that administration of CO abolished anoxia- (*in vitro*) or ischemia- (*in vivo*) induced apoptosis of lung endothelial cells in a p38 mitogen-activated protein kinase-dependent way (112). On the other hand, protection of TNF α -induced apoptosis in endothelial cells (56) by CO required basal transcription of NF α B-dependent antiapototic genes (9).

In addition, a role in angiogenesis was ascribed to HO-1 (for review, see 22). New vessel formation is important for tissue repair after acute inflammation; moreover, a relation between ongoing angiogenesis and chronic inflammatory disorders has been described (43). Recently, it has been shown *in vitro* that induction of HO-1 induces expression of vascular endothelial growth factor and promotes angiogenic activities in endothelial cells; this effect was attributed to a CO-mediated elevation of cyclic GMP (46). Furthermore, using an *in vivo* Matrigel plug assay, it was demonstrated that induction of HO-1 during angiogenesis led to a significant decrease of concomitant leukocyte infiltration (11), and it was suggested that HO-1 induction maximizes angiogenesis asso-

ciated with the resolution of tissue injury, whereas it inhibits inflammatory angiogenesis.

Taken together, products of HO-1 enzymatic activity beneficially modulate the inflammatory response toward resolution. Bilirubin seems to down-regulate inflammation in endothelial cells, whereas endothelial-derived CO exerts proresolving effects on neighboring cells, such as smooth muscle cells and leukocytes.

Nitric oxide synthase (NOS)

NOSs are oxidoreductases that convert L-arginine, tetrahy-drobiopterin (BH4), and oxygen in the presence of electron donors such as FAD, FMN, and NADPH to nitric oxide (NO) and L-citrulline. Three forms of the enzyme exist: neuronal (nNOS, NOS1), inducible (iNOS, NOS2), and endothelial NOS (eNOS, NOS3).

It is the inducible form (iNOS) that is generally controlled by inflammatory mediators. However, endothelial cell-derived NO regulates basal and stimulated vascular tone inducing vasodilation (25), displays potent antiinflammatory properties, and improves endothelial barrier function (36, 111). Mice lacking eNOS were shown to have, besides increased blood pressure (38), attenuated vascular leakage and edema formation during acute inflammation (10). Furthermore, it was shown that these mice are resistant to LPS-induced systemic hypotension (14).

Could endothelial cell-derived NO contribute to resolution of acute inflammation? In endothelial cells, NO inhibits expression of P-selectin (15), VCAM-1, ICAM-1, and Eselectin, but also of IL-6 and IL-8 by scavenging superoxide anions and thereby inhibiting the NFkB pathway (18). Furthermore, the oxidation product of NO and superoxide anion (O₂-), peroxynitrite (ONOO-), inhibits adhesion of leukocytes ex vivo (72), which is, however, dependent on its concentration (73), and attenuates rolling of neutrophils in vivo by down-regulating P-selectin (59). However, analysis of leukocyte transmigration in eNOS (-/-) mice revealed controversial results. Whereas Bucci et al. showed that in a footpad-swelling model, as well as in the air-pouch model, leukocyte traffic remained unchanged (10), Lefer et al. showed that basal leukocyte rolling, as well as thrombin-induced rolling and adherence, was greatly enhanced in eNOS (-/-) and in nNOS (-/-), but not in iNOS (-/-) mice. Furthermore, increased PMN extravasation into the peritoneal cavity after thioglycollate stimulation in eNOS (-/-) and in nNOS (-/-) mice was demonstrated (60).

An imbalance of L-arginine and BH4 leads to an uncoupling of eNOS, using oxygen as a substrate and generating superoxide (104). This would cause, on the one hand, further oxidation of BH4, perpetuating the uncoupled reaction, and, on the other hand, the formation of high concentrations of peroxynitrite by using the remaining NO. Thus, the bioavailability of NO would be reduced, and highly reactive oxygen species formed, resulting in impairment of endothelial function. Such impairment of endothelial function is commonly seen during acute inflammation and normally vanishes during resolution. Indeed, prolonged endothelial dysfunction resulting from diminished bioactivity of NO induced by oxidative stress (99), lipid overload (24), or hyperglycemia (16) has

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been implicated in the development of chronic vascular diseases. It is tempting to speculate that in these situations a dysfunctional endothelium has lost the full capacity to induce resolution of acute inflammation.

ENDOTHELIUM-DERIVED LIPID MEDIATORS IN THE RESOLUTION OF ACUTE INFLAMMATION

Phospholipase A_{γ} (PLA $_{\gamma}$)

Polyunsaturated fatty acids, such as arachidonic acid, are hydrolyzed from the *sn-2* position of membrane phospholipids by PLA₂ and can be further metabolized in endothelial cells by several enzymes releasing proinflammatory lipid mediators (74).

The three main classes of PLA₂, *i.e.*, calcium-independent PLA₂ (iPLA₂), cytosolic PLA₂ (cPLA₂), and secretory PLA₂ (sPLA₂), are further subdivided into several isoforms (55). It has been shown that these phospholipases are sequentially induced during acute inflammation; iPLA₂ was shown to mediate the onset of acute inflammation by releasing arachidonic acid for the synthesis of proinflammatory eicosanoids, as well as platelet-activating factor, whereas sPLA₂ and cPLA₂ were shown to be expressed in later phases of inflammation, promoting resolution (31).

COX-2

Inducible COX-2 is expressed at sites of inflammation. Because of the proinflammatory activity of its products, such as prostaglandin E₂ (PGE₂), and because of the beneficial effects of pharmacological inhibitors, COX-2 was generally thought to play a detrimental role. However, also antiinflammatory properties have been ascribed to COX-2 (for review, see 109). Recent data demonstrate that COX-2 is expressed during the resolution phase of inflammation and in areas of wound healing. Furthermore, clinical trials have shown that COX-2-selective inhibitors delayed the healing and tissue restoration, suggesting that COX-2 promotes resolution.

Using the carrageenan pleurisy model, Willoughby et al. showed that COX-2 and HO-1 both were essential for the resolving phase because the inhibition of these enzymes delayed the resolution of inflammation (27, 28). Further investigation revealed that the antiinflammatory effects of endothelial COX-2 during the later phase of inflammation are mediated by preferential release of prostaglandin D₂ (PGD₂) and its cyclopentenone breakdown product, 15-deoxy- Δ^{12-14} prostaglandin J₂ (15dPGJ₂), potent-Jially stimulating peroxisome proliferator-activated receptor (PPAR)-dependent transcription and other antiinflammatory mechanisms (for review, see 85). Indeed, COX-2 expression was biphasic, the first peak occurring within the first 2 h and the second, much higher peak occurring after 48 h with a concomitant shift from PGE, to 15dPGJ, production (30, 57). This late expression of COX-2 was essential for resolving inflammation, because inhibition of this second peak resulted in a prolonged inflammatory reaction (29). Moreover, COX-2 brings about resolution by inducing both PMN and macrophage apoptosis through the release of 15dPGJ₂ (30).

Taken together, endothelial-derived PGD_2 and the cyclopentenone prostaglandin $15dPGJ_2$ might contribute to the resolution of inflammation through various mechanisms that include the inhibition of proinflammatory gene expression, the induction of apoptosis, and the activation of PPARs (58).

Ligands for PPARy

The effects of PPARy ligands on the inflammatory response have been well studied, but the role of PPARy therein is still not clear. Whereas in endothelial cells the effects of 15dPGJ. are most likely independent of PPARy, PPARy-dependent effects have been shown mainly in macrophages. Moreover, whether 15dPGJ₂ is produced in vivo in sufficient amounts to activate PPARs is still a matter of debate (5). Using a highly sensitive and selective assay, Bell-Parikh et al. suggest that the amounts of 15dPGJ, generated in vivo are far too low to be compatible with a role for this substance as an endogenous activator of PPAR γ (5). Indeed, it was shown in human umbilical vein endothelial cells that 15d-PGJ, was a ligand for retinoic acid receptor-related orphan receptor-α, but not PPARγ (68). However, because of its reactivity, 15dPGJ₂, like cyclopentenone itself, can directly react with proteins and either inhibit or enhance their activity.

Recently, it was found that nitroalkene derivatives of linoleic acid (nitrolinoleic acid) are formed via NO-dependent oxidative inflammatory reactions. Nitrolinoleic acid is a significantly more robust PPAR γ ligand than other reported endogenous PPAR γ ligands, including 15dPGJ $_2$ and synthetic PPAR γ agonists such as rosiglitazone and ciglitazone. These findings show that fatty acid nitration products and PPAR-dependent gene expression can transduce NO-mediated cell signaling reactions (86), potentially down-regulating acute inflammation to induce resolution.

Ligands for PPARa

The role of ligand-induced activation of PPAR α in inflammatory control is less controversial. Activated PPARα inhibits the production of inflammatory response markers, such as endothelin-1, VCAM-1, IL-6, and tissue factor, in endothelial cells, smooth muscle cells, and macrophages (4). In endothelial cells, PPARα ligands have been found to inhibit monocyte recruitment and adhesion through down-regulation of chemoattractant and adhesion proteins (for review, see 65). PPARα reduces the activity and DNA-binding capacity of proinflammatory transcription factors such as NFkB and AP-1. This ligand-dependent transcriptional transrepression of proinflammatory genes involves competition for transcription cofactors, as well as direct interaction and interference of PPARα with p65 and c-Jun (17). However, whether the described antiinflammatory mechanisms of PPARα contribute to resolution of acute inflammation remains to be shown. A possible scenario would be the formation of oxidized fatty acids (e.g., hydroxyeicosatetraenoic acids) during neutrophildependent ROS production, which then would serve as ligands and activators of PPARα, resulting in inhibition of inflammatory gene expression, up-regulation of proresolving genes, and a switch of the inflammatory reaction. In this context, we have shown that HO-1 is a target gene for PPARα in endothelial cells (Kronke et al., manuscript submitted), further implying a role for PPARs in resolution.

Lipoxins and resolvins

In addition to prostaglandins, endothelial cells generate precursors for lipid mediators that are produced through transcellular metabolism in leukocytes and promote resolution of acute inflammation. During acute inflammation, endothelial cells together with leukocytes produce lipoxins from arachidonic acid via transcellular biosynthesis. These trihydroxytetraene-containing eicosanoids are generated through concerted activation of COX and lipoxygenases. Aspirin acetylates COX and causes the endogenous biosynthesis of so-called aspirintriggered carbon 15-epimers of lipoxins, which mimic the biologic activities of native lipoxins. These compounds serve as local endogenous antiinflammatory mediators and have been shown to trigger resolution (1, 47, 88). The major cellular targets reported for lipoxins are leukocytes, in particular neutrophils, monocytes, and eosinophils. Lipoxin A₄ (LXA₄) exhibits potent antiinflammatory properties in vitro and in animal models of acute inflammation by inhibiting neutrophil and lymphocyte activation. Therefore, LXA4 is an important endogenous counterregulatory signal that promotes resolution of acute inflammation. Moreover, it has been shown that aspirin-triggered 15-epi-LXA, and lipoxin B, analogues inhibit neutrophil-mediated changes in vascular permeability (90) and rapidly stimulate nonphlogistic phagocytosis of apoptotic neutrophils by monocyte-derived macrophages (32).

 LXA_4 and 15-epi- LXA_4 are agonists for the LXA_4 receptor, ALX. Recent studies showed that overexpression of ALX in transgenic mice resulted in accelerated resolution of induced inflammation (19, 62). Activation of ALX was shown to regulate leukocyte motility; however, in endothelial cells, where ALX is not expressed, lipoxins may also interact with cysteinyl leukotriene 1 (CysLT1) receptors (34, 98). Therefore, LXA_4 interacts with at least two classes of cell-surface receptors: ALX, specific for LXA_4 on leukocytes, and CysLT1, shared by leukotriene D_4 (LTD_4) on endothelial cells.

On the other hand, endothelial cells produce from docosahexaenoic acid, an omega-3 polyunsaturated fatty acid, metabolites such as 13-hydroxydocosahexaenoic acid (13-HDHA), and if aspirin-triggered, 17*R*-HDHA. Leukocytes can then via lipoxygenase produce so-called resolvins from these precursors (87). Resolvins represent local autacoids that display potent antiinflammatory activity, promoting resolution (91). Recently, a receptor for resolvin E1 has been identified as the orphan G protein-coupled receptor ChemR23 (2).

Together, these lipid-derived "stop signals" may be involved in switching the cellular response from additional PMN recruitment toward monocytes (in a nonphlogistic fashion) that could lead to resolution of the inflammatory response and promotion of repair and healing.

Oxidized phospholipids (OxPL) and sphingosine-1-phosphate (S1P)

During inflammation, tissue damage is caused by neutrophilderived free radicals, originally designated to destroy invading pathogens. As a consequence, oxidative modification of membrane lipids occurs and results in the formation of a variety of biologically active compounds that accumulate in the inflamed tissue. These oxidized lipids induce specific cellular reactions, which profoundly modulate the inflammatory pro-

cess. The classical view of lipid oxidation products is that they can induce and propagate chronic inflammatory reactions; however, recent data show that cells and tissues respond toward these oxidatively formed stress signals also by activation of antiinflammatory, maybe even proresolving processes.

Data from our laboratory and others indicate that the formation of phospholipid oxidation products potentially contributes to the resolving process at several levels of the inflammatory cascade: OxPL were recently shown to be generated by endothelial cells upon treatment with IL-1 (97) and during apoptosis (12, 13, 39, 49). OxPL (39, 61), but also peroxidation products of arachidonic acid, such as isoprostanes (40), stimulated endothelial cells to bind selectively monocytes, but not neutrophils (39, 61) via expression of connecting segment 1-fibronectin (92). Moreover, OxPL induce maturation of monocytes into macrophages via generation of granulocyte-macrophage colony stimulating factor (80). Furthermore, OxPL induce expression of proresolving genes such as HO-1 (42, 48, 54), COX-2 (79), and IL-10 (103), represent recognition signals for phagocytosis (12, 50), and inhibit acute inflammation induced by LPS (7).

Finally, we have recently shown that OxPL induced a sustained barrier-protective effect, counteracting thrombin-induced endothelial cell barrier disruption. We demonstrated that the effects of OxPL were additive to those of S1P (6). S1P, a biologically active lipid generated by hydrolysis of membrane lipids in activated platelets and various cell types, was demonstrated to be the major barrier-protective product of platelets (21, 84). Additionally, it has been shown *in vitro* that S1P decreases neutrophil chemotaxis and transendothelial migration in response to IL-8 (51). More recently, it was described in *in vivo* models that S1P prevents edema formation and significantly reduces neutrophil accumulation after LPS stimulation (66, 77). Thus, S1P may act in concert with OxPL to restore endothelial barrier function during acute inflammation, an important process for successful resolution.

CONCLUSION AND OUTLOOK

Resolution of acute inflammation involves highly coordinated active mechanisms that need to be switched on at a certain time point in the course of the inflammatory response. Endothelial cells play a major role in resolution, because they are able to secrete a vast array of pro- and antiinflammatory mediators that help to coordinate leukocyte traffic and barrier function. However, prolonged endothelial activation can lead to overwhelming and uncontrolled inflammation that is detrimental and can result in chronic inflammation. Therefore, a tightly balanced inflammatory response followed by timely resolution is essential for successful tissue regeneration. Further research is needed to elucidate the exact endothelial mechanisms that are involved in resolution of acute inflammation, eventually making this process a target for pharmacological intervention.

ABBREVIATIONS

ALX, lipoxin A₄ receptor; Ang-1, angiopoietin-1; AOC3, amine oxidase, copper-containing-3; AP-1, activator protein-

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1; BH4, tetrahydrobiopterin; CO, carbon monoxide; COX-2, cyclooxygenase-2; cPLA2, cytosolic PLA2; CuZn-SOD, copper, zinc superoxide dismutase; CysLT1, cysteinyl leukotriene 1; 15dPGJ_2 , $15\text{-deoxy-}\Delta^{12-14}$ -prostaglandin J_2 ; eNOS, NOS3, endothelial NOS; HDHA, hydroxydocosahexaenoic acid; HO-1, heme oxygenase-1; ICAM-1, intercellular adhesion molecule-1; IL, interleukin; iNOS, NOS2, inducible NOS: iPLA, calcium-independent PLA, LPS, lipopolysaccharide; LSP-1, leukocyte-specific protein 1; LXA₄, lipoxin A₄; NFκB, nuclear factor-κB; nNOS, NOS1, neuronal NOS; NO, nitric oxide; NOS, nitric oxide synthase; OxPL, oxidized phospholipids; PGD₂, prostaglandin D₂; PGE₂, prostaglandin E₂; PLA₂, phospholipase A₂; PMN, polymorphonuclear leukocyte; PPAR, peroxisome proliferator-activated receptor; ROS, reactive oxygen species; SOD, superoxide dismutase; S1P, sphingosine-1-phosphate; sPLA₂, secretory PLA₂; TNF α , tumor necrosis factor α ; VCAM-1, vascular cell adhesion molecule-1.

REFERENCES

- Ariel A, Chiang N, Arita M, Petasis NA, and Serhan CN. Aspirin-triggered lipoxin A4 and B4 analogs block extracellular signal-regulated kinase-dependent TNF-alpha secretion from human T cells. *J Immunol* 170: 6266–6272, 2003.
- Arita M, Bianchini F, Aliberti J, Sher A, Chiang N, Hong S, Yang R, Petasis NA, and Serhan CN. Stereochemical assignment, antiinflammatory properties, and receptor for the omega-3 lipid mediator resolvin E1. *J Exp Med* 201: 713–722, 2005.
- Arruda MA, Rossi AG, de Freitas MS, Barja-Fidalgo C, and Graca-Souza AV. Heme inhibits human neutrophil apoptosis: involvement of phosphoinositide 3-kinase, MAPK, and NF-kappaB. *J Immunol* 173: 2023–2030, 2004.
- Barbier O, Torra IP, Duguay Y, Blanquart C, Fruchart JC, Glineur C, and Staels B. Pleiotropic actions of peroxisome proliferator-activated receptors in lipid metabolism and atherosclerosis. *Arterioscler Thromb Vasc Biol* 22: 717– 726, 2002.
- 5. Bell-Parikh LC, Ide T, Lawson JA, McNamara P, Reilly M, and FitzGerald GA. Biosynthesis of 15-deoxy-delta12,14-PGJ2 and the ligation of PPARgamma. *J Clin Invest* 112: 945–955, 2003.
- Birukov KG, Bochkov VN, Birukova AA, Kawkitinarong K, Rios A, Leitner A, Verin AD, Bokoch GM, Leitinger N, and Garcia JG. Epoxycyclopentenone-containing oxidized phospholipids restore endothelial barrier function via Cdc42 and Rac. Circ Res 95: 892–901, 2004.
- Bochkov VN, Kadl A, Huber J, Gruber F, Binder BR, and Leitinger N. Protective role of phospholipid oxidation products in endotoxin-induced tissue damage. *Nature* 419: 77–81, 2002.
- 8. Brigelius-Flohe R, Banning A, and Schnurr K. Selenium-dependent enzymes in endothelial cell function. *Antioxid Redox Signal* 5: 205–215, 2003.
- Brouard S, Berberat PO, Tobiasch E, Seldon MP, Bach FH, and Soares MP. Heme oxygenase-1-derived carbon monoxide requires the activation of transcription factor

NF-kappa B to protect endothelial cells from tumor necrosis factor-alpha-mediated apoptosis. *J Biol Chem* 277: 17950–17961, 2002.

- Bucci M, Roviezzo F, Posadas I, Yu J, Parente L, Sessa WC, Ignarro LJ, and Cirino G. Endothelial nitric oxide synthase activation is critical for vascular leakage during acute inflammation in vivo. *Proc Natl Acad Sci U S A* 102: 904–908, 2005.
- Bussolati B, Ahmed A, Pemberton H, Landis RC, Di CF, Haskard DO, and Mason JC. Bifunctional role for VEGFinduced heme oxygenase-1 in vivo: induction of angiogenesis and inhibition of leukocytic infiltration. *Blood* 103: 761–766, 2004.
- 12. Chang MK, Bergmark C, Laurila A, Horkko S, Han KH, Friedman P, Dennis EA, and Witztum JL. Monoclonal antibodies against oxidized low-density lipoprotein bind to apoptotic cells and inhibit their phagocytosis by elicited macrophages: evidence that oxidation-specific epitopes mediate macrophage recognition. *Proc Natl Acad Sci U S A* 96: 6353–6358, 1999.
- Chang MK, Binder CJ, Miller YI, Subbanagounder G, Silverman GJ, Berliner JA, and Witztum JL. Apoptotic cells with oxidation-specific epitopes are immunogenic and proinflammatory. *J Exp Med* 200: 1359–1370, 2004.
- Connelly L, Madhani M, and Hobbs AJ. Resistance to endotoxic shock in endothelial nitric-oxide synthase (eNOS) knock-out mice: a pro-inflammatory role for eNOS-derived NO in vivo. J Biol Chem 280: 10040–10046, 2005.
- Davenpeck KL, Gauthier TW, and Lefer AM. Inhibition of endothelial-derived nitric oxide promotes P-selectin expression and actions in the rat microcirculation. *Gastroen*terology 107: 1050–1058, 1994.
- Davi G, Falco A, and Patrono C. Lipid peroxidation in diabetes mellitus. Antioxid Redox Signal 7: 256–268, 2005.
- Daynes RA and Jones DC. Emerging roles of PPARs in inflammation and immunity. *Nat Rev Immunol* 2: 748–759, 2002.
- 18. De Caterina R, Libby P, Peng HB, Thannickal VJ, Rajavashisth TB, Gimbrone MA Jr, Shin WS, and Liao JK. Nitric oxide decreases cytokine-induced endothelial activation. Nitric oxide selectively reduces endothelial expression of adhesion molecules and proinflammatory cytokines. *J Clin Invest* 96: 60–68, 1995.
- Devchand PR, Arita M, Hong S, Bannenberg G, Moussignac RL, Gronert K, and Serhan CN. Human ALX receptor regulates neutrophil recruitment in transgenic mice: roles in inflammation and host defense. *FASEB J* 17: 652–659, 2003.
- Didion SP, Kinzenbaw DA, Fegan PE, Didion LA, and Faraci FM. Overexpression of CuZn-SOD prevents lipopolysaccharide-induced endothelial dysfunction. *Stroke* 35: 1963–1967, 2004.
- Dudek SM and Garcia JG. Cytoskeletal regulation of pulmonary vascular permeability. J Appl Physiol 91: 1487–1500, 2001.
- 22. Dulak J, Jozkowicz A, Foresti R, Kasza A, Frick M, Huk I, Green CJ, Pachinger O, Weidinger F, and Motterlini R. Heme oxygenase activity modulates vascular endothelial growth factor synthesis in vascular smooth muscle cells. *Antioxid Redox Signal* 4: 229–240, 2002.

- Eltzschig HK, Ibla JC, Furuta GT, Leonard MO, Jacobson KA, Enjyoji K, Robson SC, and Colgan SP. Coordinated adenine nucleotide phosphohydrolysis and nucleoside signaling in posthypoxic endothelium: role of ectonucleotidases and adenosine A2B receptors. *J Exp Med* 198: 783–796, 2003.
- Esenabhalu VE, Schaeffer G, and Graier WF. Free fatty acid overload attenuates Ca²⁺ signaling and NO production in endothelial cells. *Antioxid Redox Signal* 5: 147–153, 2003
- Furchgott RF and Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* 288: 373–376, 1980.
- Gamble JR, Drew J, Trezise L, Underwood A, Parsons M, Kasminkas L, Rudge J, Yancopoulos G, and Vadas MA. Angiopoietin-1 is an antipermeability and anti-inflammatory agent in vitro and targets cell junctions. *Circ Res* 87: 603–607, 2000.
- Gilroy DW, Tomlinson A, and Willoughby DA. Differential effects of inhibition of isoforms of cyclooxygenase (COX-1, COX-2) in chronic inflammation. *Inflamm Res* 47: 79–85, 1998.
- Gilroy DW, Tomlinson A, and Willoughby DA. Differential effects of inhibitors of cyclooxygenase (cyclooxygenase 1 and cyclooxygenase 2) in acute inflammation. Eur J Pharmacol 355: 211–217, 1998.
- 29. Gilroy DW, Colville-Nash PR, Willis D, Chivers J, Paul-Clark MJ, and Willoughby DA. Inducible cyclooxygenase may have anti-inflammatory properties. *Nat Med* 5: 698–701, 1999.
- Gilroy DW, Colville-Nash PR, McMaster S, Sawatzky DA, Willoughby DA, and Lawrence T. Inducible cyclooxygenase-derived 15-deoxy(Delta)12–14PGJ2 brings about acute inflammatory resolution in rat pleurisy by inducing neutrophil and macrophage apoptosis. *FASEB J* 17: 2269–2271, 2003.
- Gilroy DW, Newson J, Sawmynaden P, Willoughby DA, and Croxtall JD. A novel role for phospholipase A2 isoforms in the checkpoint control of acute inflammation. *FASEB J* 18: 489–498, 2004.
- Godson C, Mitchell S, Harvey K, Petasis NA, Hogg N, and Brady HR. Cutting edge: lipoxins rapidly stimulate nonphlogistic phagocytosis of apoptotic neutrophils by monocytederived macrophages. *J Immunol* 164: 1663–1667, 2000.
- Goepfert C, Imai M, Brouard S, Csizmadia E, Kaczmarek E, and Robson SC. CD39 modulates endothelial cell activation and apoptosis. *Mol Med* 6: 591–603, 2000.
- 34. Gronert K, Martinsson-Niskanen T, Ravasi S, Chiang N, and Serhan CN. Selectivity of recombinant human leukotriene D(4), leukotriene B(4), and lipoxin A(4) receptors with aspirin-triggered 15-epi-LXA(4) and regulation of vascular and inflammatory responses. *Am J Pathol* 158: 3–9, 2001.
- Guckelberger O, Sun XF, Sevigny J, Imai M, Kaczmarek E, Enjyoji K, Kruskal JB, and Robson SC. Beneficial effects of CD39/ecto-nucleoside triphosphate diphosphohydrolase-1 in murine intestinal ischemia–reperfusion injury. *Thromb Haemost* 91: 576–586, 2004.
- 36. Gupta MP, Ober MD, Patterson C, Al-Hassani M, Natarajan V, and Hart CM. Nitric oxide attenuates H₂O₂-induced

- endothelial barrier dysfunction: mechanisms of protection. *Am J Physiol Lung Cell Mol Physiol* 280: L116–L126, 2001.
- 37. Hayashi S, Takamiya R, Yamaguchi T, Matsumoto K, Tojo SJ, Tamatani T, Kitajima M, Makino N, Ishimura Y, and Suematsu M. Induction of heme oxygenase-1 suppresses venular leukocyte adhesion elicited by oxidative stress: role of bilirubin generated by the enzyme. *Circ Res* 85: 663–671, 1999.
- Huang PL, Huang Z, Mashimo H, Bloch KD, Moskowitz MA, Bevan JA, and Fishman MC. Hypertension in mice lacking the gene for endothelial nitric oxide synthase. *Nature* 377: 239–242, 1995.
- Huber J, Vales A, Mitulovic G, Blumer M, Schmid R, Witztum JL, Binder BR, and Leitinger N. Oxidized membrane vesicles and blebs from apoptotic cells contain biologically active oxidized phospholipids that induce monocyte–endothelial interactions. *Arterioscler Thromb Vasc Biol* 22: 101–107, 2002.
- Huber J, Bochkov VN, Binder BR, and Leitinger N. The isoprostane 8-iso-PGE2 stimulates endothelial cells to bind monocytes via cyclic AMP- and p38 MAP kinasedependent signaling pathways. *Antioxid Redox Signal* 5: 163–169, 2003.
- Imai M, Goepfert C, Kaczmarek E, and Robson SC. CD39 modulates IL-1 release from activated endothelial cells. *Biochem Biophys Res Commun* 270: 272–278, 2000.
- 42. Ishikawa K, Navab M, Leitinger N, Fogelman AM, and Lusis AJ. Induction of heme oxygenase-1 inhibits the monocyte transmigration induced by mildly oxidized LDL. *J Clin Invest* 100: 1209–1216, 1997.
- 43. Jackson JR, Seed MP, Kircher CH, Willoughby DA, and Winkler JD. The codependence of angiogenesis and chronic inflammation. *FASEB J* 11: 457–465, 1997.
- 44. Jamieson T, Cook DN, Nibbs RJ, Rot A, Nixon C, McLean P, Alcami A, Lira SA, Wiekowski M, and Graham GJ. The chemokine receptor D6 limits the inflammatory response in vivo. *Nat Immunol* 6: 403–411, 2005.
- Jongstra-Bilen J, Misener VL, Wang C, Ginzberg H, Auerbach A, Joyner AL, Downey GP, and Jongstra J. LSP1 modulates leukocyte populations in resting and inflamed peritoneum. *Blood* 96: 1827–1835, 2000.
- Jozkowicz A, Huk I, Nigisch A, Weigel G, Dietrich W, Motterlini R, and Dulak J. Heme oxygenase and angiogenic activity of endothelial cells: stimulation by carbon monoxide and inhibition by tin protoporphyrin-IX. Antioxid Redox Signal 5: 155–162, 2003.
- 47. Jozsef L, Zouki C, Petasis NA, Serhan CN, and Filep JG. Lipoxin A4 and aspirin-triggered 15-epi-lipoxin A4 inhibit peroxynitrite formation, NF-kappa B and AP-1 activation, and IL-8 gene expression in human leukocytes. *Proc Natl Acad Sci U S A* 99: 13266–13271, 2002.
- 48. Kadl A, Huber J, Gruber F, Bochkov VN, Binder BR, and Leitinger N. Analysis of inflammatory gene induction by oxidized phospholipids in vivo by quantitative real-time RT-PCR in comparison with effects of LPS. *Vascul Phar-macol* 38: 219–227, 2002.
- Kadl A, Bochkov VN, Huber J, and Leitinger N. Apoptotic cells as sources for biologically active oxidized phospholipids. *Antioxid Redox Signal* 6: 311–320, 2004.

1752 KADL AND LEITINGER

50. Kagan VE, Borisenko GG, Serinkan BF, Tyurina YY, Tyurin VA, Jiang J, Liu SX, Shvedova AA, Fabisiak JP, Uthaisang W, and Fadeel B. Appetizing rancidity of apoptotic cells for macrophages: oxidation, externalization, and recognition of phosphatidylserine. *Am J Physiol Lung Cell Mol Physiol* 285: L1–L17, 2003.

- Kawa S, Kimura S, Hakomori S, and Igarashi Y. Inhibition of chemotactic motility and trans-endothelial migration of human neutrophils by sphingosine 1-phosphate. *FEBS Lett* 420: 196–200, 1997.
- Kawamura K, Ishikawa K, Wada Y, Kimura S, Matsumoto H, Kohro T, Itabe H, Kodama T, and Maruyama Y. Bilirubin from heme oxygenase-1 attenuates vascular endothelial activation and dysfunction. *Arterioscler Thromb Vasc Biol* 25: 155–160, 2005.
- 53. Koskinen K, Vainio PJ, Smith DJ, Pihlavisto M, Yla-Herttuala S, Jalkanen S, and Salmi M. Granulocyte transmigration through the endothelium is regulated by the oxidase activity of vascular adhesion protein-1 (VAP-1). *Blood* 103: 3388–3395, 2004.
- 54. Kronke G, Bochkov VN, Huber J, Gruber F, Bluml S, Furnkranz A, Kadl A, Binder BR, and Leitinger N. Oxidized phospholipids induce expression of human heme oxygenase-1 involving activation of cAMP-responsive element-binding protein. *J Biol Chem* 278: 51006–51014, 2003
- Kudo I and Murakami M. Phospholipase A2 enzymes. Prostaglandins Other Lipid Mediat 68–69: 3–58, 2002.
- Kushida T, Li VG, Quan S, Goodman A, and Abraham NG. Role of human heme oxygenase-1 in attenuating TNFalpha-mediated inflammation injury in endothelial cells. J Cell Biochem 87: 377–385, 2002.
- Lawrence T, Gilroy DW, Colville-Nash PR, and Willoughby DA. Possible new role for NF-kappaB in the resolution of inflammation. *Nat Med* 7: 1291–1297, 2001.
- Lawrence T, Willoughby DA, and Gilroy DW. Anti-inflammatory lipid mediators and insights into the resolution of inflammation. *Nat Rev Immunol* 2: 787–795, 2002.
- Lefer DJ, Scalia R, Campbell B, Nossuli T, Hayward R, Salamon M, Grayson J, and Lefer AM. Peroxynitrite inhibits leukocyte–endothelial cell interactions and protects against ischemia–reperfusion injury in rats. *J Clin Invest* 99: 684–691, 1997.
- Lefer DJ, Jones SP, Girod WG, Baines A, Grisham MB, Cockrell AS, Huang PL, and Scalia R. Leukocyte–endothelial cell interactions in nitric oxide synthase-deficient mice. *Am J Physiol* 276: H1943–H1950, 1999.
- 61. Leitinger N, Tyner TR, Oslund L, Rizza C, Subbanagounder G, Lee H, Shih PT, Mackman N, Tigyi G, Territo MC, Berliner JA, and Vora DK. Structurally similar oxidized phospholipids differentially regulate endothelial binding of monocytes and neutrophils. *Proc Natl Acad Sci U S A* 96: 12010–12015, 1999.
- Levy BD, De Sanctis GT, Devchand PR, Kim E, Ackerman K, Schmidt BA, Szczeklik W, Drazen JM, and Serhan CN. Multi-pronged inhibition of airway hyper-responsiveness and inflammation by lipoxin A(4). *Nat Med* 8: 1018–1023, 2002
- 63. Lin SJ, Shyue SK, Hung YY, Chen YH, Ku HH, Chen JW, Tam KB, and Chen YL. Superoxide dismutase inhibits the expression of vascular cell adhesion molecule-1 and intra-

- cellular cell adhesion molecule-1 induced by tumor necrosis factor-alpha in human endothelial cells through the JNK/p38 pathways. *Arterioscler Thromb Vasc Biol* 25: 334–340, 2005.
- 64. Liu L, Cara DC, Kaur J, Raharjo E, Mullaly SC, Jongstra-Bilen J, Jongstra J, and Kubes P. LSP1 is an endothelial gatekeeper of leukocyte transendothelial migration. *J Exp Med* 201: 409–418, 2005.
- 65. Marx N, Duez H, Fruchart JC, and Staels B. Peroxisome proliferator-activated receptors and atherogenesis: regulators of gene expression in vascular cells. *Circ Res* 94: 1168–1178, 2004.
- 66. McVerry BJ, Peng X, Hassoun PM, Sammani S, Simon BA, and Garcia JG. Sphingosine 1-phosphate reduces vascular leak in murine and canine models of acute lung injury. *Am J Respir Crit Care Med* 170: 987–993, 2004.
- Middleton J, Patterson AM, Gardner L, Schmutz C, and Ashton BA. Leukocyte extravasation: chemokine transport and presentation by the endothelium. *Blood* 100: 3853– 3860, 2002.
- 68. Migita H and Morser J. 15-Deoxy-Δ¹²,¹⁴-prostaglandin J² (15d-PGJ2) signals through retinoic acid receptor-related orphan receptor-alpha but not peroxisome proliferator-activated receptor-gamma in human vascular endothelial cells: the effect of 15d-PGJ2 on tumor necrosis factor-alpha-induced gene expression. Arterioscler Thromb Vasc Biol 25: 710–716, 2005.
- Mizumoto N, Kumamoto T, Robson SC, Sevigny J, Matsue H, Enjyoji K, and Takashima A. CD39 is the dominant Langerhans cell-associated ecto-NTPDase: modulatory roles in inflammation and immune responsiveness. *Nat Med* 8: 358–365, 2002.
- Moncure M, Chen L, Childs EW, Smalley D, Udobi KF, and Cheung LY. Heme-oxygenase-1 mRNA expression affects hemorrhagic shock-induced leukocyte adherence. *J Trauma* 55: 118–125, 2003.
- Morse D, Pischke SE, Zhou Z, Davis RJ, Flavell RA, Loop T, Otterbein SL, Otterbein LE, and Choi AM. Suppression of inflammatory cytokine production by carbon monoxide involves the JNK pathway and AP-1. *J Biol Chem* 278: 36993–36998, 2003.
- Nossuli TO, Hayward R, Scalia R, and Lefer AM. Peroxynitrite reduces myocardial infarct size and preserves coronary endothelium after ischemia and reperfusion in cats. *Circulation* 96: 2317–2324, 1997.
- Nossuli TO, Hayward R, Jensen D, Scalia R, and Lefer AM. Mechanisms of cardioprotection by peroxynitrite in myocardial ischemia and reperfusion injury. *Am J Physiol* 275: H509–H519, 1998.
- O'Donnell VB. Free radicals and lipid signaling in endothelial cells. *Antioxid Redox Signal* 5: 195–203, 2003.
- Otterbein LE. Carbon monoxide: innovative anti-inflammatory properties of an age-old gas molecule. *Antioxid Redox Signal* 4: 309–319, 2002.
- Otterbein LE, Bach FH, Alam J, Soares M, Tao LH, Wysk M, Davis RJ, Flavell RA, and Choi AM. Carbon monoxide has anti-inflammatory effects involving the mitogenactivated protein kinase pathway. *Nat Med* 6: 422–428, 2000.
- 77. Peng X, Hassoun PM, Sammani S, McVerry BJ, Burne MJ, Rabb H, Pearse D, Tuder RM, and Garcia JG. Protective

- effects of sphingosine 1-phosphate in murine endotoxininduced inflammatory lung injury. *Am J Respir Crit Care Med* 169: 1245–1251, 2004.
- Pizurki L, Zhou Z, Glynos K, Roussos C, and Papapetropoulos A. Angiopoietin-1 inhibits endothelial permeability, neutrophil adherence and IL-8 production. *Br J Pharmacol* 139: 329–336, 2003.
- Pontsler AV, St Hilaire A, Marathe GK, Zimmerman GA, and McIntyre TM. Cyclooxygenase-2 is induced in monocytes by peroxisome proliferator activated receptor gamma and oxidized alkyl phospholipids from oxidized low density lipoprotein. *J Biol Chem* 277: 13029–13036, 2002.
- Rajavashisth TB, Andalibi A, Territo MC, Berliner JA, Navab M, Fogelman AM, and Lusis AJ. Induction of endothelial cell expression of granulocyte and macrophage colony-stimulating factors by modified low-density lipoproteins. *Nature* 344: 254–257, 1990.
- Rueckschloss U, Duerrschmidt N, and Morawietz H. NADPH oxidase in endothelial cells: impact on atherosclerosis. *Antioxid Redox Signal* 5: 171–180, 2003.
- Savill J. Apoptosis in resolution of inflammation. J Leukoc Biol 61: 375–380, 1997.
- Savill J and Haslett C. Granulocyte clearance by apoptosis in the resolution of inflammation. *Semin Cell Biol* 6: 385–393, 1995.
- 84. Schaphorst KL, Chiang E, Jacobs KN, Zaiman A, Natarajan V, Wigley F, and Garcia JG. Role of sphingosine-1 phosphate in the enhancement of endothelial barrier integrity by platelet-released products. *Am J Physiol Lung Cell Mol Physiol* 285: L258–L267, 2003.
- Scher JU and Pillinger MH. 15d-PGJ2: the anti-inflammatory prostaglandin? Clin Immunol 114: 100–109, 2005.
- Schopfer FJ, Lin Y, Baker PR, Cui T, Garcia-Barrio M, Zhang J, Chen K, Chen YE, and Freeman BA. Nitrolinoleic acid: an endogenous peroxisome proliferator-activated receptor gamma ligand. *Proc Natl Acad Sci U S A* 102: 2340–2345, 2005.
- Serhan CN. A search for endogenous mechanisms of antiinflammation uncovers novel chemical mediators: missing links to resolution. *Histochem Cell Biol* 122: 305–321, 2004.
- Serhan CN and Chiang N. Lipid-derived mediators in endogenous anti-inflammation and resolution: lipoxins and aspirin-triggered 15-epi-lipoxins. *ScientificWorldJournal* 2: 169–204, 2002.
- Serhan CN and Levy B. Novel pathways and endogenous mediators in anti-inflammation and resolution. *Chem Immunol Allergy* 83: 115–145, 2003.
- Serhan CN, Takano T, Clish CB, Gronert K, and Petasis N. Aspirin-triggered 15-epi-lipoxin A4 and novel lipoxin B4 stable analogs inhibit neutrophil-mediated changes in vascular permeability. Adv Exp Med Biol 469: 287–293, 1999.
- 91. Serhan CN, Gotlinger K, Hong S, and Arita M. Resolvins, docosatrienes, and neuroprotectins, novel omega-3-derived mediators, and their aspirin-triggered endogenous epimers: an overview of their protective roles in catabasis. *Prostaglandins Other Lipid Mediat* 73: 155–172, 2004.
- 92. Shih PT, Elices MJ, Fang ZT, Ugarova TP, Strahl D, Territo MC, Frank JS, Kovach NL, Cabanas C, Berliner JA, and Vora DK. Minimally modified low-density lipoprotein induces monocyte adhesion to endothelial connecting seg-

- ment-1 by activating beta1 integrin. *J Clin Invest* 103: 613–625, 1999.
- Soares MP, Usheva A, Brouard S, Berberat PO, Gunther L, Tobiasch E, and Bach FH. Modulation of endothelial cell apoptosis by heme oxygenase-1-derived carbon monoxide. *Antioxid Redox Signal* 4: 321–329, 2002.
- 94. Soares MP, Seldon MP, Gregoire IP, Vassilevskaia T, Berberat PO, Yu J, Tsui TY, and Bach FH. Heme oxygenase-1 modulates the expression of adhesion molecules associated with endothelial cell activation. *J Immunol* 172: 3553–3563, 2004.
- Stolen CM, Marttila-Ichihara F, Koskinen K, Yegutkin GG, Turja R, Bono P, Skurnik M, Hanninen A, Jalkanen S, and Salmi M. Absence of the endothelial oxidase AOC3 leads to abnormal leukocyte traffic in vivo. *Immunity* 22: 105–115, 2005.
- 96. Stone PC, Lally F, Rahman M, Smith E, Buckley CD, Nash GB, and Rainger GE. Transmigrated neutrophils down-regulate the expression of VCAM-1 on endothelial cells and inhibit the adhesion of flowing lymphocytes. *J Leukoc Biol* 77: 44–51, 2005.
- 97. Subbanagounder G, Wong JW, Lee H, Faull KF, Miller E, Witztum JL, and Berliner JA. Epoxyisoprostane and epoxycyclopentenone phospholipids regulate monocyte chemotactic protein-1 and interleukin-8 synthesis. Formation of these oxidized phospholipids in response to interleukin-1beta. *J Biol Chem* 277: 7271–7281, 2002.
- Takano T, Fiore S, Maddox JF, Brady HR, Petasis NA, and Serhan CN. Aspirin-triggered 15-epi-lipoxin A4 (LXA4) and LXA4 stable analogues are potent inhibitors of acute inflammation: evidence for anti-inflammatory receptors. *J Exp Med* 185: 1693–1704, 1997.
- Thomas SR, Chen K, and Keaney JF Jr. Oxidative stress and endothelial nitric oxide bioactivity. *Antioxid Redox Signal* 5: 181–194, 2003.
- 100. Thurston G, Suri C, Smith K, McClain J, Sato TN, Yancopoulos GD, and McDonald DM. Leakage-resistant blood vessels in mice transgenically overexpressing angiopoietin-1. *Science* 286: 2511–2514, 1999.
- 101. Thurston G, Rudge JS, Ioffe E, Zhou H, Ross L, Croll SD, Glazer N, Holash J, McDonald DM, and Yancopoulos GD. Angiopoietin-1 protects the adult vasculature against plasma leakage. *Nat Med* 6: 460–463, 2000.
- 102. Tohka S, Laukkanen M, Jalkanen S, and Salmi M. Vascular adhesion protein 1 (VAP-1) functions as a molecular brake during granulocyte rolling and mediates recruitment in vivo. FASEB J 15: 373–382, 2001.
- 103. Varadhachary AS, Monestier M, and Salgame P. Reciprocal induction of IL-10 and IL-12 from macrophages by low-density lipoprotein and its oxidized forms. *Cell Immunol* 213: 45–51, 2001.
- 104. Vasquez-Vivar J, Kalyanaraman B, Martasek P, Hogg N, Masters BS, Karoui H, Tordo P, and Pritchard KA Jr. Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors. *Proc Natl Acad Sci U S A* 95: 9220–9225, 1998.
- 105. Wakai A, Wang JH, Winter DC, Street JT, O'Sullivan RG, and Redmond HP. Adenosine inhibits neutrophil vascular endothelial growth factor release and transendothelial migration via A2B receptor activation. *Shock* 15: 297–301, 2001.

1754 KADL AND LEITINGER

106. Wang C, Hayashi H, Harrison R, Chiu B, Chan JR, Ostergaard HL, Inman RD, Jongstra J, Cybulsky MI, and Jongstra-Bilen J. Modulation of Mac-1 (CD11b/CD18)-mediated adhesion by the leukocyte-specific protein 1 is key to its role in neutrophil polarization and chemotaxis. *J Immunol* 169: 415–423, 2002.

- 107. Willis D, Moore AR, Frederick R, and Willoughby DA. Heme oxygenase: a novel target for the modulation of the inflammatory response. *Nat Med* 2: 87–90, 1996
- 108. Willis D, Moore AR, and Willoughby DA. Heme oxygenase isoform expression in cellular and antibodymediated models of acute inflammation in the rat. J Pathol 190: 627–634, 2000.
- Willoughby DA, Moore AR, Colville-Nash PR, and Gilroy D. Resolution of inflammation. *Int J Immunophar-macol* 22: 1131–1135, 2000.
- Witzenbichler B, Westermann D, Knueppel S, Schultheiss HP, and Tschope C. Protective role of angiopoietin-1 in endotoxic shock. *Circulation* 111: 97– 105, 2005.

- 111. Wong D, Dorovini-Zis K, and Vincent SR. Cytokines, nitric oxide, and cGMP modulate the permeability of an in vitro model of the human blood–brain barrier. *Exp Neurol* 190: 446–455, 2004.
- 112. Zhang X, Shan P, Otterbein LE, Alam J, Flavell RA, Davis RJ, Choi AM, and Lee PJ. Carbon monoxide inhibition of apoptosis during ischemia—reperfusion lung injury is dependent on the p38 mitogen-activated protein kinase pathway and involves caspase 3. *J Biol Chem* 278: 1248–1258, 2003.

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- 6. Alexandra Kadl, Elena Galkina, Norbert Leitinger. 2009. Induction of CCR2-dependent macrophage accumulation by oxidized phospholipids in the air-pouch model of inflammation. *Arthritis & Rheumatism* **60**:5, 1362-1371. [CrossRef]
- 7. Shana Marmon, Joseph Hinchey, Philmo Oh, Michael Cammer, Cecilia J. de Almeida, Leslie Gunther, Cedric S. Raine, Michael P. Lisanti. 2009. Caveolin-1 Expression Determines the Route of Neutrophil Extravasation through Skin Microvasculature. *The American Journal of Pathology* **174**:2, 684-692. [CrossRef]
- 8. S Vuletic, BA Taylor, GH Tofler, A Chait, SM Marcovina, K Schenck, JJ Albers. 2008. SAA and PLTP activity in plasma of periodontal patients before and after full-mouth tooth extraction. *Oral Diseases* 14:6, 514-519. [CrossRef]
- 9. Christopher P. Palmer, Maria E. Mycielska, Hakan Burcu, Kareem Osman, Timothy Collins, Rachel Beckerman, Rebecca Perrett, Helen Johnson, Ebru Aydar, Mustafa B. A. Djamgoz. 2008. Single cell adhesion measuring apparatus (SCAMA): application to cancer cell lines of different metastatic potential and voltage-gated Na+ channel expression. *European Biophysics Journal* 37:4, 359-368. [CrossRef]
- 10. Weiguo Hu, Sean P Ferris, Rodney K Tweten, Gongxiong Wu, Svetlana Radaeva, Bin Gao, Roderick T Bronson, Jose A Halperin, Xuebin Qin. 2008. Rapid conditional targeted ablation of cells expressing human CD59 in transgenic mice by intermedilysin. *Nature Medicine* **14**:1, 98-103. [CrossRef]
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- 13. Giuseppe Malleo, Emanuela Mazzon, Tiziana Genovese, Rosanna Di Paola, Carmelo Mui??, Tommaso Centorrino, Ajith K. Siriwardena, Salvatore Cuzzocrea. 2007. ETANERCEPT ATTENUATES THE DEVELOPMENT OF CERULEIN-INDUCED ACUTE PANCREATITIS IN MICE. *Shock* 27:5, 542-551. [CrossRef]